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# Convergent synthesis of an octasaccharide fragment of the O-specific polysaccharide of *Shigella dysenteriae* type 1 \*

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#### Abstract

A stereocontrolled, convergent synthesis is described of the linear octasaccharide methyl glycoside  $\alpha$ -L-Rha p- $(1 \rightarrow 2)$ - $\alpha$ -D-Gal p- $(1 \rightarrow 3)$ - $\alpha$ -Glc pNAc- $(1 \rightarrow 3)$ - $\alpha$ -L-Rha p- $(1 \rightarrow 2)$ - $\alpha$ -D-Gal p- $(1 \rightarrow 3)$ - $\alpha$ -D-Glc pNAc- $(1 \rightarrow 3)$ - $\alpha$ -L-Rha p-OMe (11), which corresponds to two contiguous repeating units of the O-specific polysaccharide of *Shigella dysenteriae* type 1.

# 1. Introduction

Robbins and co-workers [2,3] suggested, that conjugate vaccines consisting of the O-specific polysaccharides (O-SP) of *Shigellae* and an immunogenic protein would confer protective immunity to humans against shigellosis. We assumed [1] that oligosaccharides, shorter than the native polysaccharide, may also be exploited to elicit high-avidity antibodies against the O-SP of *Sh. dysenteriae* type 1, provided that they have a high degree of conformational similarity to the native O-SP. Synthetic carbohydrate chemistry may provide suitable haptens, and the purpose of the present work is to further explore the feasibility of this approach [1,4–7].

The O-specific polysaccharide of Sh. dysenteriae type 1, is a heteropolysaccharide, characterized by the tetrasaccharide repeating-unit [8,9] I. The constituent

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- 1  $\alpha$ -L-Rha p-(1  $\rightarrow$  3)- $\alpha$ -L-Rha p-OMe
- 2  $\alpha$ -D-Glc pNHCOC<sub>2</sub>H<sub>5</sub>-(1  $\rightarrow$  3)- $\alpha$ -L-Rha p-OMe
- 3  $\alpha$ -L-Rha p-(1  $\rightarrow$  2)- $\alpha$ -D-Gal p-OMe
- 4  $\alpha$ -D-Gal p- $(1 \rightarrow 3)$ - $\alpha$ -D-Glc pNAc-OMe
- 5  $\alpha$ -D-Gal p- $(1 \rightarrow 3)$ - $\alpha$ -D-Glc p NAc- $(1 \rightarrow 3)$ - $\alpha$ -L-Rha p-OMe
- 6 α-L-Rha p-(1 → 2)-α-D-Gal p-(1 → 3)-α-D-Glc pNAc-(1 → 3) -α-L-Rha p-OMe
- 7  $\alpha$ -D-Gal p- $(1 \rightarrow 3)$ - $\alpha$ -D-Glc p NAc- $(1 \rightarrow 3)$ - $\alpha$ -L-Rha p- $(1 \rightarrow 3)$ - $\alpha$ -L-Rha p-OMe
- 8  $\alpha$ -D-Gal p- $(1 \rightarrow 3)$ - $\alpha$ -D-Glc p NHCOC  $_2$  H  $_5$ - $(1 \rightarrow 3)$ - $\alpha$ -L-Rha p- $(1 \rightarrow 3)$ - $\alpha$ -L-Rha p-OMe
- 9  $\alpha$ -L-Rha p- $(1 \rightarrow 3)$ - $\alpha$ -L-Rha p- $(1 \rightarrow 2)$ - $\alpha$ -D-Gal p- $(1 \rightarrow 3)$ - $\alpha$ -D-Glc pNAc- $(1 \rightarrow 3)$ - $\alpha$ -L-Rha p-OMe
- 10  $\alpha$ -L-Rha p- $(1 \rightarrow 3)$ - $\alpha$ -L-Rha p- $(1 \rightarrow 2)$ - $\alpha$ -D-Gal p- $(1 \rightarrow 3)$   $-\alpha$ -D-Glc pNAc- $(1 \rightarrow 3)$ - $\alpha$ -L-Rha p- $(1 \rightarrow 3)$ - $\alpha$ -L-Rha p-OMe

monosaccharide units are  $\alpha$ -linked D-galactose, 2-acetamido-2-deoxy-D-glucose, and L-rhamnose. Hitherto we have prepared [1,4,5,10] the methyl glycosides of dito hexa-saccharide fragments (1-10) of the O-SP. The use of these oligosaccharides as molecular probes in mapping the binding characteristics of a murine monoclonal antibody against the O-SP has been reported [11]. We have also described the synthesis of two fully protected, frame-shifted tetrasaccharides related to the O-specific polysaccharide, that can function both as glycosyl donors, and as glycosyl acceptors, after chemoselective removal of only one of their protecting groups [6,7]. These intermediates are now being used in our laboratory as building blocks for at the synthesis of extended epitopes of the O-SP. We are employing high resolution NMR spectroscopy to characterize the synthetic oligosaccharides and to estimate their similarity to the O-SP. To this end, we completely assigned the 500 or 600 MHz <sup>1</sup>H, and 100 MHz <sup>13</sup>C NMR spectra of oligosaccharides 1-10, which have the natural,  $\alpha$  anomeric configuration at their reducing end termini [4,5,10]. A comparison of selected NMR characteristics of oligosaccharides 1-10 with those of the O-SP led us to conclude, that short oligosaccharides fail to mimic the conformational features of the native polysaccharide [4,5]. They are, therefore, unlikely to elicit protective antibodies. Even the

hexasaccharide 10 lacks a high degree of conformational similarity to the O-SP [5]. A more complex fragment is therefore needed for a conformational mimicry. Here we describe a convergent synthesis of the homologous octasaccharide methyl glycoside 11, which corresponds to two contiguous repeating units of the O-SP.

## 2. Results and discussion

Experience gained during the preparation of the smaller homologues 1-10 indicated that the construction of the trans interglycosidic linkage between the two rhamnose residues (units **D** and **E**) is more convenient than that between any other two units in 11. We based our strategy on condensation of two tetrasaccharide building blocks corresponding to the ABCD and EFGH segments. These blocks were constructed in a stepwise manner. The ABCD segment was prepared as follows. The disaccharide [4] 12 was converted to the acetamido derivative 13 by reduction of the azido group, followed by N-acetylation [12], in 79% (Scheme 1). Reaction of the glycosyl acceptor 13 with methyl 3,4,6-tri-O-benzyl-2-O-(4methoxybenzyl)-1-thio- $\beta$ -p-galactopyranoside [4.5] (14) under promotion by methyl trifluoromethanesulfonate [13] (MeOTf) afforded the ABC trisaccharide 15 in 86% yield (Scheme 2). The  $\alpha$  stereochemistry of the newly formed interglycosidic linkage was inferred from the <sup>3</sup>J<sub>H-1,H-2</sub> coupling constant for the galactosyl residue, which is 3.8 Hz. The glycosylation reaction was accompanied by partial anomerization of the thio-galactoside donor to 16, which we noted and characterized earlier [4]. Next, the oxidative removal of the 4-methoxybenzyl group from 15 with 2,3-dichloro-5,6-dicyano-1,4-benzoquinone [14] (DDO) afforded the trisaccharide acceptor 17 in 57% yield. The strategy for the rhamnose unit D envisaged a temporary, selectively removable protecting group at O-3, and permanent groups

\*Reagents: (a) NiCl2 . 6 H<sub>2</sub>O, H<sub>3</sub>BO<sub>3</sub>, NaBH<sub>4</sub> — (MeOCH<sub>2</sub>)<sub>2</sub>, EtOH; (b) Ac<sub>2</sub>O — MeOH.

#### Scheme 1.

at O-2 and O-4. Compound 29 was selected for this purpose, in which the bromoacetyl group [15] fulfills our requirement, since it can be removed selectively, under essentially neutral conditions. We designed several approaches to this unit. In the first route, the starting compound was methyl  $\alpha$ -L-rhamnopyranoside 18, which was converted to the dibenzoate 21 in a three-step, one-pot reaction through the intermediacy of the cyclic orthoesters 19 and 20 (Scheme 3, reactions a and b). The combined yield from this protocol is 60%. The route follows the method of Garegg and Hultberg [16], which was adapted for 21 by Wessel and Bundle [17]. Reproduction of the published protocol [17] led to several experimental improvements, which made the method suitable also for the high-yielding syntheses of 2-O-mono- and 2,4-di-O-acyl-1-thio-rhamnosides [7,18]. As a result of these developments, 21 could now be obtained as a crystalline substance. Alternatively, 21 was

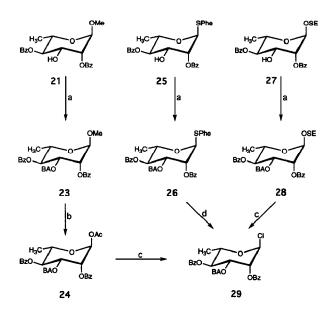
\*Reagents: (a)  $CF_3SO_3Me$ , 2,6-di-tert-butyl-4-methyl- $C_5H_2N$  —  $(C_2H_5)_2O$ ; (b) 2,3-dichloro-5,6-dicyano-1,4-benzoquinone —  $CH_2Cl_2$ ,  $H_2O$ .

MB = 4-methoxybenzyl

\*Reagents: (a) PhC(OMe)<sub>3</sub>, H<sup>+</sup>; (b) BzCl —  $C_5H_5N$ ; (c) AcOH —  $H_2O$ ; (d) (CH<sub>3</sub>)<sub>2</sub>C(OCH<sub>3</sub>)<sub>2</sub>, 10-camphorsulfonic acid; (e) BzCl —  $C_5H_5N$ .

### Scheme 3.

obtained from 18 through the diol [19] 22 in a combined yield of 66%. Bromoacetylation [15] of 21 afforded 23 [20] which was acetolyzed (H<sub>2</sub>SO<sub>4</sub>-Ac<sub>2</sub>O) to give the acetate 24 (72% yield) (Scheme 4).



 $^{\mathrm{e}}\mathrm{Reagents:} \text{ (a) } \mathrm{BrCH_{2}COBr} - \mathrm{C}_{5}\mathrm{H}_{5}\mathrm{N}; \text{ (b) } \mathrm{Ac}_{2}\mathrm{O}, \mathrm{H}_{2}\mathrm{SO}_{4}; \text{ (c) } \mathrm{CH}_{3}\mathrm{OCHCl}_{2}, \mathrm{ZnCl}_{2} \cdot \mathrm{Et}_{2}\mathrm{O} - \mathrm{CH}_{2}\mathrm{Cl}_{2}; \\$ 

(d)  $Cl_2 - CH_2Cl_2$ .

BA = bromoacetyl SE = 2-(trimethylsilyl)ethyl

Scheme 4.

\*Reagents: (a)  $CF_3SO_3Ag$ , 2,6-di-tert-butyl-4-methyl- $C_5H_2N$  —  $CH_2Cl_2$ ; (b)  $CS(NH_2)_2$  — MeOH. BA = bromoacetyl

Scheme 5.

Treatment [21] of 24 with 1,1-dichloromethyl methyl ether and zinc chloride—diethyl ether complex [4] afforded the rhamnosyl chloride 29 in a nearly quantitative yield. The  $\alpha$  anomeric configuration in 29 was ascertained by the 183 Hz  $^1J_{\text{C-1,H-1}}$  coupling constant [22]. Alternative routes to 29 included the conversion of the known phenyl 1-thiorhamnoside [18] 25 and 2-(trimethylsilyl)ethyl rhamnoside [7] 27 into their O-bromoacetyled derivatives 26 and 28, respectively, which were routinely transformed to 29 as shown in Scheme 4. Condensation of the ABC acceptor 17 with the rhamnosyl donor 29 in the presence of silver trifluoromethanesulfonate (AgOTf) and 2,6-di-tert-butyl-4-methylpyridine (DTBMP) gave the fully protected ABCD tetrasaccharide 30 in an excellent yield. Removal of the bromoacetyl group with thiourea [15] afforded the tetrasaccharide aceptor 31 in 94% yield (Scheme 5).

We chose the trisaccharide (trimethylsilyl)ethyl glycoside [6] **32** as the precursor to the **EFGH** block. 2-(Trimethylsilyl)ethyl glycosides of mono- and oligo-saccharides have been shown to be useful intermediates in oligosaccharide syntheses [23–25]. Whereas their glycosidic linkage is stable under a variety of reaction conditions, they can be converted to glycosyl donors in high yields either directly,

\*Reagents: (a) 2,3-dichloro-5,6-dicyano-1,4-benzoquinone — CH<sub>2</sub>Cl<sub>2</sub>, H<sub>2</sub>O.

MB = 4-methoxybenzyl SE = 2-(trimethylsilyl)ethyl

\*Reagents: (a)  $CF_3SO_3Ag$ , 2,6-di-tert-butyl-4-methyl- $C_5H_2N$  —  $CH_2Cl_2$ ; (b) AcOH,  $H_2O$ ; (c)  $Ac_2O$  —  $C_5H_5N$ ; (d)  $CF_3COOH$  —  $CH_2Cl_2$ ; (e)  $CCl_3CN$ , 1,8-diazabicyclo[5.4.0]undec-7-ene —  $CH_2Cl_2$ .

SE = 2-(trimethylsilyl)ethyl

#### Scheme 7.

or by way of the corresponding glycosyl hemiacetals [24]. Treatment of the compound 32 with DDO afforded the alcohol 33 in 94% yield (Scheme 6). The trisaccharide acceptor 33 was glycosylated with 2,3,4-tri-O-benzoyl- $\alpha$ -L-rhamnopyranosyl bromide [26] 34 in the presence of AgOTf and DTBMP, to give the protected tetrasaccharide 35 in 94% yield (Scheme 7). We found, that the 34 or the corresponding chloride [7] are less likely to promote side-reactions, e.g., formation of an orthoester or a  $\beta$ -rhamnosyl linkage, than the corresponding O-acetylated counterparts. Next, the benzylidene-acetal function was replaced by acetyl groups [(i) H<sub>3</sub>O<sup>+</sup>, (ii) Ac<sub>2</sub>O-Py] to afford 36. An attempt to convert the tetrasaccharide glycoside 36 into the corresponding tetraosyl chloride proved to be abortive under the conditions (1,1-dichloromethyl methyl ether, ZnCl<sub>2</sub>) reported for related transformations [23,24]. Although the 2-(trimethylsilyl)ethoxy group in 36 could be replaced by chlorine during this reaction, concurrent cleavage of residue H occurred, as shown by the isolation of 2,3,4-tri-O-benzoyl- $\alpha$ -L-rhamnopyranosyl chloride [7] in pure form from the mixture. An approximately equimolar amount of a glycotriosyl chloride was also isolated, which corresponds to the EFG sequence. An alternative approach was therefore sought for the transformation of 36 into a glycosyl donor. This was accomplished in a two-step sequence. First, the 2-(trimethylsilyl)ethyl group was removed by the treatment [23] of 36 with trifluoroacetic acid in CH<sub>2</sub>Cl<sub>2</sub>, to afford the hemiacetal 37 in 87% yield. Subsequently, compound 37 was converted to the trichloroacetimidate derivative 38 by reaction [27] with

trichloroacetonitrile in the presence of 1,8-diazabicyclo[5.4.0]undec-7-ene [28] in 94% yield. The  $\alpha$  anomeric configuration of the reducing end unit in 38 was inferred from the chemical shift analogy of H-1<sub>E</sub> with the corresponding proton in a closely related compound [6].

Condensation of the tetrasaccharide donor 38 with the tetrasaccharide acceptor 17 was performed in CH<sub>2</sub>Cl<sub>2</sub>, under catalysis by boron trifluoride etherate. The fully protected octasaccharide 39 could be isolated in 69% yield. The stereochemical integrity of 39 was indicated by NMR spectroscopy, which provided proof for the all- $\alpha$  stereochemistry of the interglycosidic linkages. The identity of compound 39 received further support from fast-atom bombardment mass spectroscopy (FABMS), using a mixture of dithiothreitol and dithioerythritol as the matrix. For 39 the measured mass of the protonated parent ion was 2994.028 amu. This is within 11 ppm from the calculated mass for C<sub>167</sub>H<sub>165</sub>N<sub>4</sub>O<sub>48</sub> which is 2994.059. To achieve this accuracy, data from several scans were acquired in profile mode which allows averaging of peak shape before assignment of mass. A two-step conversion of 39 [(i) NiCl<sub>2</sub>-H<sub>3</sub>BO<sub>3</sub>-NaBH<sub>4</sub>; (ii) Ac<sub>2</sub>O] afforded the acetamido derivative 40 in 44% yield. The measured mass of the protonated parent ion was 3010.059, which agrees within 7 ppm with the theoretical value calculated for  $C_{169}H_{169}N_2O_{49}$ . Routine deprotection of 40 [(i) NaOMe-MeOH, (ii) H<sub>2</sub>-Pd-C] afforded the octasaccharide methyl glycoside 11, which was characterized by NMR and FABMS. At this stage, partial assignment of the <sup>1</sup>H NMR spectrum of 11 was aided by the published spectra of the smaller homologues 1-10. We compared the chemical shifts of the anomeric protons for 11 with those in the O-SP. The criterion for coincidence was set arbitrarily as less than 0.01 ppm difference in the chemical shifts [5]. This comparison revealed, that in 11 the chemical shifts of the anomeric protons of five consecutive residues coincide with the corresponding chemical shifts for the O-SP. These are the residues C, D, E, F, and G. It is very likely that increasing spectral coincidence reflects increasing conformational similarities. Thus we believe, the octasaccharide 11 more closely approaches the conformational determinant of the O-SP than any of the smaller homologues 1–10. In order to further probe the similarity between the octasaccharide 11 and the O-SP of Sh. dysenteriae type 1, a detailed analysis of their <sup>1</sup>H and <sup>13</sup>C NMR spectra is currently being carried out in our laboratory. These findings will be the subject of a future publication [29].

# 3. Experimental

General methods.—General experimental conditions are described in ref. 4. Optical rotations were measured for CHCl<sub>3</sub> solutions, except where indicated otherwise. The NMR data were obtained on a Gemini 300 (Varian) spectrometer, operating at 300 MHz for <sup>1</sup>H and at 75 MHz for <sup>13</sup>C. The fast-atom bombardment mass spectra were run on a Jeol SX102 mass spectrometer using 6 keV Xe atoms to ionize the samples which were desorbed from a "magic bullet" matrix (mixture of dithiothreitol and dithioerythritol). For low mass spectra (< 2000 amu), the instrument was calibrated against Ultramark 1621 (PCR Chemicals) and for higher mass samples, against cesium iodide. For the low resolution, chemical ionization mass spectra (CIMS) ammonia was used as the ionizing gas.

Methyl O-(2-acetamido-4,6-O-benzylidene-2-deoxy- $\alpha$ -D-glucopyranosyl)-(1  $\rightarrow$  3)-2,4-di-O-benzyl-α-D-rhamnopyranoside (13).—To a solution of methyl O-(2-azido-4,6-O-benzylidene-2-deoxy- $\alpha$ -D-glucopyranosyl)- $(1 \rightarrow 3)$ -2,4-di-O-benzyl- $\alpha$ -L-rhamnopyranoside [4] (12, 2.9 g, 4.5 mmol) in EtOH (20 mL) was added a solution of NiCl<sub>2</sub>·6H<sub>2</sub>O (3.0 g, 12.6 mmol) and H<sub>3</sub>BO<sub>3</sub> (1.8 g, 29 mmol) in EtOH (120 mL). To this solution was added, under stirring at 25°C, a 1% solution of NaBH<sub>4</sub> in EtOH (~40 mL) during 1 h. The mixture was cooled to 0°C, then treated with Ac<sub>2</sub>O (2 mL). Extractive work-up, followed by column chromatography using EtOAc as eluant afforded 13 as an amorphous solid (2.35 g, 79%);  $[\alpha]_D + 15^\circ$  (c 0.5). NMR (CDCl<sub>3</sub>):  ${}^{1}$ H,  $\delta$  7.15–7.45 (aromatic), 5.519 (s, 1 H, CHPh),  $\bar{4}$ .932 (d, 1 H,  $J_{1,2}$  3.9 Hz, H-1<sub>B</sub>), 4.771, 4.744, 4.586, and 4.507 (4 d, 4 H, 2 C $H_2$  of Bn), 3.337 (s, 3 H, C $H_3$ O), 1.618 (s, 3 H, C $H_3$ CO), and 1.373 (d, 1 H,  $J_{5,6}$  6.2 Hz, H-6<sub>A</sub>); <sup>13</sup>C, δ 172.1 (C=O), 137.4–126.5 (aromatic), 101.9 (CHPh), 97.5 (C-1<sub>A</sub>), 93.6 (C-1<sub>B</sub>), 82.1 (C-4<sub>B</sub>), 75.6 and 71.9 (CH<sub>2</sub> of Bn), 68.6 (C-6<sub>B</sub>), 54.9 (CH<sub>3</sub>O), 54.3 (C-2<sub>B</sub>), 22.6 (CH<sub>3</sub>CO), and 18.1 (C-6<sub>A</sub>). CIMS: m/z 650 [(M + H)<sup>+</sup>]. Anal. Calcd for C<sub>36</sub>H<sub>43</sub>NO<sub>10</sub>: C, 66.54; H, 6.67; N, 2.16. Found: C, 66.45; H, 6.71; N, 2.13.

Methyl O-[3,4,6-tri-O-benzyl-2-O-(4-methoxybenzyl)- $\alpha$ -D-galactopyranosyl]-(1  $\rightarrow$  3)-O-(2-acetamido-4,6-O-benzylidene-2-deoxy- $\alpha$ -D-glucopyranosyl)-(1  $\rightarrow$  3)-2,4-di-O-benzyl- $\alpha$ -L-rhamnopyranoside (15).—Methyl trifluoromethanesulfonate (150  $\mu$ L) was added to a stirred mixture of 13 (2.92 g, 4.5 mmol), methyl 3,4,6-tri-O-benzyl-2-O-(4-methoxybenzyl)-1-thio- $\beta$ -D-galactopyranoside [4,5] (14, 5.0 g, 8.3 mmol), 2,6-di-tert-butyl-4-methylpyridine (1.5 g, 7.4 mmol), and 4A molecular sieves (3 g)

in diethyl ether (100 mL), at 25°C. The mixture was stirred for 60 h, during which time more MeOTf was added after 20 h (150  $\mu$ L, 1.3 mmol), and 40 h (200  $\mu$ L, 1.8 mmol). The reaction was terminated by the addition of Et<sub>3</sub>N (3 mL). Work-up in the usual manner followed by chromatography (3:1  $\rightarrow$ 2:1 hexane–EtOAc) afforded first a mixture of 14 and 16 (Ref 4), from which 14 (1.5 g) was isolated by crystallization. Subsequent elution afforded 15 as an amorphous solid (4.70 g, 87%);  $[\alpha]_D$  +48° (c 0.8). NMR (CDCl<sub>3</sub>):  $^1$ H,  $\delta$  7.4–6.63 (aromatic), 6.382 (d, 1 H, NH), 5.545 (d, 1 H,  $J_{1,2}$  3.8 Hz, H-1<sub>C</sub>), 5.417 (s, 1 H, CHPh), 5.082 (d, 1 H,  $J_{1,2}$  3.7 Hz, H-1<sub>B</sub>), 3.746 (s, 3 H, CH<sub>3</sub>O of 4-methoxybenzyl), 3.279 [s, 3 H, CH<sub>3</sub>O (aglycon)], 1.701 (CH<sub>3</sub>CO), 1.355 (d, 1 H,  $J_{5,6}$  6.2 Hz, H-6<sub>A</sub>);  $^{13}$ C,  $\delta$  158.9–113.4 (aromatic), 101.8 (CHPh), 98.1 and 97.1 (C-1<sub>A</sub>, 1<sub>C</sub>), 93.0 (C-1<sub>B</sub>), 82.8 (C-4<sub>B</sub>), 75.8, 74.3, 73.8, 73.7, 72.3, 71.0, 70.7, and 68.8 (C-6<sub>B</sub>, 6<sub>C</sub>, and CH<sub>2</sub> of Bn and 4-methoxybenzyl), 55.1 and 54.7 (CH<sub>3</sub>O), 51.8 (C-2<sub>B</sub>), 22.7 (CH<sub>3</sub>CO), and 17.9 (C-6<sub>A</sub>). FABMS: m/z 1202 [(M + H)<sup>+</sup>]. Anal. Calcd for C<sub>71</sub>H<sub>79</sub>NO<sub>16</sub>: C, 70.92; H, 6.62; N, 1.16. Found: C, 71.01; H, 6.91; N, 1.18.

Methyl O-(3,4,6-tri-O-benzyl-α-D-galactopyranosyl)-(1 → 3)-O-(2-acetamido-4,6-O-benzylidene-2-deoxy-α-D-glucopyranosyl)-(1 → 3)-2,4-di-O-benzyl-α-L-rhamnopyranoside (17).—2,3-Dichloro-5,6-dicyano-1,4-benzoquinone (0.8 g, 3.5 mmol) was added to a stirred mixture of 15 (3.3 g, 2.7 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (200 mL) and H<sub>2</sub>O (10 mL) at 25°C. After 2 h, more 2,3-dichloro-5,6-dicyano-1,4-benzoquinone (0.4 g, 1.8 mmol) was added, and the mixture was stirred for an additional period of 2 h. The usual work-up, followed by chromatography (1:1 hexane-EtOAc) gave 17 as an amorphous substance (1.7 g, 57%);  $[\alpha]_D$  +43° (c 0.7). NMR (CDCl<sub>3</sub>): <sup>1</sup>H, δ 5.543 (s, 1 H, CHPh), 5.350 (d, 1 H,  $J_{1,2}$  4.2 Hz, H-1<sub>C</sub>), 5.022 (m, 1 H, H-1<sub>B</sub>), 3.295 (s, 3 H, CH<sub>3</sub>O), and 1.375 (d, 1 H,  $J_{5,6}$  6.3 Hz, H-6<sub>A</sub>); <sup>13</sup>C, δ 172.2 (C=O), 138.4-126.0 (aromatic), 101.1 (CHPh), 99.9 (C-1<sub>A</sub>), 97.9 (C-1<sub>C</sub>), 93.6 (C-1<sub>B</sub>), 82.3 (C-4<sub>B</sub>), 75.9, 74.5, 73.7, 73.2, and 72.3 (CH<sub>2</sub> of Bn), 68.8 and 68.7 (C-6<sub>A</sub>,6<sub>B</sub>), 55.8 (CH<sub>3</sub>O), 52.1 (C-2<sub>B</sub>), 22.9 (CH<sub>3</sub>CO), and 18.2 (C-6<sub>A</sub>). FABMS: m/z 1082 [(M+H)<sup>+</sup>]. Anal. Calcd for C<sub>63</sub>H<sub>71</sub>NO<sub>15</sub>: C, 69.91; H, 6.61; N, 1.29. Found: C, 69.70; H, 6.94; N, 1.26.

Methyl 4-O-benzoyl-α-L-rhamnopyranoside (22).—To a solution of methyl 2,3-O-isopropylidene-α-L-rhamnopyranoside, obtained from 18 (5 g, 28 mmol) as described in ref 30, in pyridine (8 mL) was added BzCl (3.6 mL) at 0°C. After 1 h, the mixture was concentrated. Extractive work-up (CHCl<sub>3</sub>-5% aq HCl-H<sub>2</sub>O) afforded a semicrystalline product, which was dissolved in 4:1 MeOH-H<sub>2</sub>O (25 mL). To this mixture was added CF<sub>3</sub>CO<sub>2</sub>H (2 mL). After 30 min, the volatiles were evaporated under vacuum. The residue was dried by repeated addition and evaporation of toluene. Treatment of the syrupy residue with petroleum ether afforded crystalline 22 (5.7 g, 72%); mp 113-115°C;  $[\alpha]_D - 106^\circ$  (c 1.1), lit. [19]  $[\alpha]_D - 82^\circ$  (CHCl<sub>3</sub>). NMR (CDCl<sub>3</sub>):  $^1$ H, δ 8.06-7.36 (aromatic), 5.090 (t, 1 H,  $J_{3,4} = J_{4,5} = 9.4$  Hz, H-4), 4.753 (br d, 1 H, H-1), 4.04-3.98 (m, 2 H, H-2,3), 3.931 (dq, 1 H, H-5), 3.398 (s, 3 H, CH<sub>3</sub>O), and 1.275 (d, 3 H,  $J_{5,6}$  6.2 Hz, H-6);  $^{13}$ C, δ 167.3 (C=O), 133.4-128.4 (aromatic), 100.6 (C-1), 75.8 (C-4), 70.8 and 70.2 (C-2,3), 65.6 (C-5), 55.0 (CH<sub>3</sub>O), and 17.4 (C-6). CIMS: m/z 300 [(M + NH<sub>4</sub>)+]. Anal. Calcd for C<sub>14</sub>H<sub>18</sub>O<sub>6</sub>: C, 59.57; H, 6.43. Found: C, 59.40; H, 6.50.

Methyl 2.4-di-O-benzoyl- $\alpha$ -L-rhamnopyranoside (21).—(a) A mixture of 18 (25 g, 88 mmol), trimethyl orthobenzoate (43 mL, 250 mmol), and a catalytic amount of 10-camphorsulfonic acid was stirred at 25°C until the mixture became homogeneous (~30 min), then the by-product MeOH was removed at 30°C, under the vacuum of a water aspirator. The residue, containing the intermediate 19, was dissolved in pyridine (50 mL). To this solution was added BzCl (24 mL), at 0°C. After 3 h, the mixture was treated with aq NaHCO3 at 0°C, then extracted with CHCl<sub>3</sub>. The organic phase was washed with H<sub>2</sub>O, then concentrated to give a syrup, containing the intermediate 20. Residual pyridine was removed by repeated addition and evaporation of H<sub>2</sub>O. To a solution of the syrupy residue in CH<sub>3</sub>CN (200 mL) was added aq 50% CF<sub>3</sub>CO<sub>2</sub>H (10 mL) at 0°C. After 3 min, the volatiles were removed under vacuum. Column chromatography (6:1 hexane-EtOAc) of the residue afforded a syrup which was crystallized from hexane to afford crystalline 21 (32.5 g, 60%); mp 84-86°C;  $[\alpha]_D$  +64° (c 1.3), lit. [17]  $[\alpha]_D$  +57° (CHCl<sub>3</sub>), lit. [31] for the D isomer,  $[\alpha]_D - 59^\circ$  (CHCl<sub>3</sub>). CIMS: m/z 404 [(M +  $NH_4$ )<sup>+</sup>], 387 [(M + H)<sup>+</sup>], and 355 [(M + H - MeOH)<sup>+</sup>]. Anal. Calcd for  $C_{21}H_{22}O_7$ : C, 65.27; H, 5.75. Found: C, 65.30; H, 5.78.

(b) To a mixture of 22 (5.0 g, 17.6 mmol) and trimethyl orthobenzoate (6 mL, 35 mmol) was added a catalytic amount of 10-camphorsulfonic acid. The mixture was shaken until it became homogeneous ( $\sim 10$  min). The by-product MeOH was removed as described in (a). To a solution of the intermediate orthoester 20 in CH<sub>3</sub>CN (50 mL) at 0°C was added aq 50% CF<sub>3</sub>CO<sub>2</sub>H (2 mL). After 3 min, the mixture was processed as described in (a), to give 21 (6.3 g, 92%), which was identical to the product obtained in (a).

1-O-Acetyl-2,4-di-O-benzoyl-3-O-bromoacetyl-α-L-rhamnopyranose (24).—To a solution of methyl 2,4-di-O-benzoyl-3-O-bromoacetyl-α-L-rhamnopyranoside [20] (23, 5.5 g, 11 mmol), obtained from 21 by bromoacetylation essentially as described [15] in Ac<sub>2</sub>O (40 mL) at 0°C was added conc H<sub>2</sub>SO<sub>4</sub> (6 drops). After 3 h, solid NaHCO<sub>3</sub> was added, and the mixture was stirred for 5 min. The mixture was concentrated. Extractive work-up, followed by column chromatography using 5:1 hexane–EtOAc as eluant gave amorphous 24 (4.2 g, 72%);  $[\alpha]_D$  +60° (c 1.2). NMR (CDCl<sub>3</sub>): <sup>1</sup>H, δ 8.13–7.43 (aromatic), 6.238 (d, 1 H,  $J_{1,2}$  1.9 Hz, H-1), 5.645 (dd, 1 H,  $J_{2,3}$  3.6,  $J_{3,4}$  10.2 Hz, H-3), 5.572 (dd, 1 H, H-2), 5.551 (t, 1 H, H-4), 4.177 (dq, 1 H, H-5), 3.670 and 3.620 (2 d, 2 H, J ~ 11.9 Hz, C $H_2$ Br), 2.236 (s, 3 H, C $H_3$ CO), 1.355 (d, 3 H,  $J_{5,6}$  6.3 Hz, H-6); <sup>13</sup>C, δ 169.0 (C=O of Ac), 167.4 (C=O of BrCH<sub>2</sub>CO), and 166.1 (2 C) (C=O of Bz), 133.9–128.6 (aromatic), 90.7 ( $^1J_{C-1,H-1}$  177 Hz, C-1), 70.8, 70.7, and 69.1 (2 C) (C-2,3,4,5), 24.9 (CH<sub>2</sub>Br), 20.9 (CH<sub>3</sub>CO), and 17.6 (C-6). CIMS: m/z 552 ([M + NH<sub>4</sub>)<sup>+</sup>] and 475 [(M + H – AcOH)<sup>+</sup>]. Anal. Calcd for C<sub>24</sub>H<sub>23</sub>BrO<sub>9</sub>: C, 53.84; H, 4.33; Br, 14.93. Found: C, 54.10; H, 4.55; Br, 14.79.

Phenyl 2,4-di-O-benzoyl-3-O-bromoacetyl-1-thio-α-L-rhamnopyranoside (26).— Phenyl 2,4-di-O-benzoyl-1-thio-α-L-rhamnopyranoside [18] (25) was bromoacetylated as described for the preparation of compound 23, to afford 26 as a syrup (94%);  $[\alpha]_D - 28^\circ$  (c 1.4). NMR (CDCl<sub>3</sub>): <sup>1</sup>H, δ 8.12–7.24 (aromatic), 5.816 (dd, 1 H,  $J_{1,2}$  1.7,  $J_{2,3}$  2.9 Hz, H-2), 5.625–5.551 (m, 3 H, H-1,3,4), 4.602 (dq, 1 H, H-5),

3.667 and 3.625 (2 d, 2 H,  $CH_2Br$ ), and 1.360 (d, 1 H,  $J_{5,6}$  6.2 Hz, H-6);  $^{13}C$ ,  $\delta$  166.4, 165.54, and 165.48 (C=O), 133.6–128.0 (aromatic), 85.8 (C-1), 71.8, 71.5, and 71.4 (C-2,3,4), 68.1 (C-5), 25.0 ( $CH_2Br$ ) and 17.6 (C-6). CIMS: m/z 602 [( $M+NH_4$ )+]. Anal. Calcd for  $C_{28}H_{25}BrO_7S$ : C, 57.44; H, 4.30; Br, 13.65; S, 5.48. Found: C, 56.59; C, 439; C, 58, 5.33.

- 2-(Trimethylsilyl)ethyl 2,4-di-O-benzoyl-3-O-bromoacetyl-α-L-rhamnopyranoside (28).—2-(Trimethylsilyl)ethyl 2,4-di-O-benzoyl-α-L-rhamnopyranoside [7] (27) was bromoacetylated as described for the preparation of 23, to afford 28 as a syrup (71%);  $[\alpha]_D$  +52° (c 1). NMR (CDCl<sub>3</sub>):  $^1$ H, δ 8.14–7.42 (aromatic), 5.658 (dd, 1 H,  $J_{2,3}$  3.4,  $J_{3,4}$  10.1 Hz, H-3), 5.516 (dd, 1 H, H-2), 5.503 (t, 1 H,  $J_{3,4} = J_{4,5} = 10.1$  Hz, H-4), 4.970 (d, 1 H, H-1), 4.123 (dq, 1 H, H-5), 3.92–3.85 and 3.66–3.57 (m, 2 H, OC  $H_2$ ), 3.652 and 3.601 (2 d, 2 H, C $H_2$ Br), and 1.336 (d, 1 H,  $J_{5,6}$  6.2 Hz, H-6);  $^{13}$ C, δ 166.5, 165.72, and 165.46 (C=O), 133.5–128.5 (aromatic), 100.0 (C-1), 71.6, 71.3, 70.6, 66.6, and 65.8 (C-2,3,4,5,OC  $H_2$ ), 68.1 (C-5), 25.1 (CH $_2$ Br), 17.9 (CH $_2$ Si), 17.6 (C-6), and 1.4 (SiMe<sub>3</sub>). CIMS: m/z 610 [(M + NH $_4$ )+]. Anal. Calcd for C $_{27}$ H $_{33}$ BrO $_8$ Si: C, 54.63; H, 5.60; Br, 13.46. Found: C, 54.53; H, 5.64; Br, 13.53.
- 2,4-DiO-benzoyl-3-O-bromoacetyl-α-L-rhamnopyranosyl chloride (29).—(a) To a stirred mixture of 24 (3.2 g, 6.0 mmol) and ZnCl<sub>2</sub>· Et<sub>2</sub>O (1 mL of a 54% solution in CH<sub>2</sub>Cl<sub>2</sub>) in CH<sub>2</sub>Cl<sub>2</sub> was added 1,1-dichloromethyl methyl ether (1.5 mL, 16.5 mmol). After 4 h aq NaHCO<sub>3</sub> was added. The CHCl<sub>3</sub> phase was washed with H<sub>2</sub>O and concentrated. Column chromatographic purification (8:1 hexane–EtOAc) of the residue afforded 29 as a syrup (3.0 g, 98%);  $[\alpha]_D$  +47°(c 1.6), lit. [20] +43°. NMR (CDCl<sub>3</sub>): <sup>1</sup>H, δ 8.13–7.42 (aromatic), 6.160 (d, 1 H, H-1), 5.907 (dd, 1 H,  $J_{2,3}$  3.4,  $J_{3,4}$  10.2 Hz, H-3), 5.701 (dd, 1 H,  $J_{1,2}$  1.7 Hz, H-2), 5.558 (t, 1 H, H-4), 4.402 (dq, 1 H, H-5), 3.655 and 3.603 (2 d, 2 H, CH<sub>2</sub>Br), and 1.385 (d, 1 H,  $J_{5,6}$  6.2 Hz, H-6); <sup>13</sup>C, δ 166.4, 165.5, and 165.2 (C=O), 133.9–128.5 (aromatic), 89.0 ( $J_{C-1,H-1}$  183 Hz, C-1), 72.3 (C-2), 70.7 (C-4), and 69.7 (C-3, 5), 24.8 (CH<sub>2</sub>Br), and 17.3 (C-6). Anal. Calcd for C<sub>12</sub>H<sub>17</sub>ClO<sub>7</sub>: C, 46.68 H, 5.55; Cl, 11.48. Found: C, 46.58; H, 5.60; Cl, 11.57.
- (b) To a solution of 26 (280 mg) in  $CH_2Cl_2$  (6 mL) was added a solution of  $Cl_2$  in  $CCl_4$  until the yellow color persisted, at 0°C. After 5 h, hex-1-ene (excess) was added and the solution concentrated. Column chromatography (8:1 hexane–EtOAc) afforded 29 as a syrup (226 mg, 92%) which was identical to the preparation obtained in (a).
- (c) Treatment of 28 with 1,1-dichloromethyl methyl ether and  $ZnCl_2 \cdot Et_2O$  as described in (a) afforded 29 (86%) which was identical to the preparation obtained in (a).

Methyl O-(2,4-di-O-benzoyl-3-O-bromoacetyl- $\alpha$ -L-rhamnopyranosyl)- $(1 \rightarrow 2)$ -O-(3,4,6-tri-O-benzyl- $\alpha$ -D-galactopyranosyl- $(1 \rightarrow 3)$ -O-(2-acetamido-4,6,-benzylidene-2-deoxy- $\alpha$ -D-glucopyranosyl)- $(1 \rightarrow 3)$ -2,4-di-O-benzyl- $\alpha$ -L-rhamnopyranoside (30).—A mixture of 17 (1.20 g, 1.1 mmol), 29 (1.14 g, 3.7 mmol), 2,6-di-tert-butyl-4-methyl-pyridine (0.4 g, 2 mmol), 4A molecular sieves (1 g), and dry CH<sub>2</sub>Cl<sub>2</sub> (50 mL) was stirred for 1 h then cooled to  $-50^{\circ}$ C. Silver trifluoromethanesulfonate (0.6 g, 2.7 mmol) was added. After 1 h more 29 (0.5 g, 1.6 mmol) and silver trifluoromethanesulfonate (0.4 g, 1.8 mmol) were added. The stirred mixture was allowed to reach

20°C in 3 h. Ice-cold, aq NaHCO<sub>3</sub> was added and the mixture was filtered. The organic phase was separated and concentrated. Column chromatography (1:1 hexane–EtOAc) of the residue afforded **30** as an amorphous solid (1.62 g, 94%);  $[\alpha]_D + 67^\circ$  (c 0.9). NMR (CDCl<sub>3</sub>):  $^1$ H,  $\delta$  8.04–6.92 (aromatic), 6.197 (d, 1 H,  $J_{NH-H-2}$  9.6 Hz, NH), 5.741 (dd, 1 H,  $J_{2,3}$  3.3,  $J_{3,4}$  10.3 Hz, H-3<sub>D</sub>), 5.707 (d, 1 H,  $J_{1,2}$  4.1 Hz, H-1<sub>C</sub>), 5.662 (dd, 1 H, H-2<sub>D</sub>), 5.657 (s, 1 H, CHPh), 5.483 (d, 1 H, H-1<sub>D</sub>), 5.250 (t, 1 H, H-4<sub>D</sub>), 5.041 (d, 1 H,  $J_{1,2}$  3.9 Hz, H-1<sub>B</sub>), 3.296 (s, 3 H, CH<sub>3</sub>O), 1.706 (s, 3 H, CH<sub>3</sub>CO), 1.340 and 0.529 (2 d, 2 H,  $J_{5,6} \sim 6$  Hz, H-6<sub>A</sub>, 6<sub>D</sub>);  $^{13}$ C,  $\delta$  169.5 (C=O of Ac), 166.3, 165.6, and 165.0 (C=O of Bz and BrCH<sub>2</sub>CO), 137.9–125.9 (aromatic), 100.4 (CHPh), 97.9 and 97.8 (C-1<sub>A</sub>,1<sub>D</sub>), 97.1 (C-1<sub>C</sub>), 92.9 (C-1<sub>B</sub>), 82.9 (C-4<sub>B</sub>), 75.9, 74.4, 73.7, 73.2, and 72.2 (CH<sub>2</sub> of Bn), 70.5 and 68.6 (C-6<sub>B</sub>,6<sub>C</sub>), 54.8 (CH<sub>3</sub>O), 51.6 (C-2<sub>B</sub>), 25.4 (CH<sub>2</sub>Br), 23.1 (CH<sub>3</sub>CO), 18.1 and 17.3 (C-6<sub>A</sub>,6<sub>B</sub>). FABMS: m/z 1558 [(C<sub>85</sub>H<sub>90</sub>BrNO<sub>22</sub> + H)<sup>+</sup>] and 1556 [(C<sub>85</sub>H<sub>90</sub> $r^{9}$ BrNO<sub>22</sub> + H)<sup>+</sup>]. Anal. Calcd for C<sub>85</sub>H<sub>90</sub>BrNO<sub>22</sub>: C, 65.54; H, 5.82; Br, 5.13; N, 0.90. Found: C, 64.82; H, 5.86; Br, 5.04; N, 0.84.

Methyl  $O-(2,4-di-O-benzoyl-\alpha-L-rhamnopyranosyl)-(1 \rightarrow 2)-O-(3,4,6-tri-O-benzyl \alpha$ -D-galactopyranosyl)- $(1 \rightarrow 3)$ -O-(2-acetamido-4,6-O-benzylidene-2-deoxy- $\alpha$ -D-glucopyranosyl)- $(1 \rightarrow 3)$ -2,4-di-O-benzyl- $\alpha$ -L-rhamnopyranoside (31).—Thiourea (0.5 g, 6.6 mmol) was added to a solution of 30 (1.5 g, 0.96 mmol) in 2:1 CH<sub>3</sub>OH-CH<sub>2</sub>Cl<sub>2</sub> (60 mL) at 25°C. After 1 h the solution was concentrated. The residue was treated with CHCl<sub>3</sub> (80 mL), the mixture filtered, and the insoluble part discarded. The CHCl<sub>3</sub> solution was washed with H<sub>2</sub>O, dried (Na<sub>2</sub>SO<sub>4</sub>), and concentrated. Column chromatography (1:1 hexane-EtOAc) of the residue afforded amorphous 31 (1.30 g, 94%);  $[\alpha]_D$  + 56° (c 0.4). NMR (CDCl<sub>3</sub>): <sup>1</sup>H,  $\delta$  8.04–6.97 (aromatic), 6.172 (d, 1 H,  $J_{NH-H-2}$  9.5 Hz, NH), 5.702 (d, 1 H,  $J_{1.2}$  4.1 Hz, H-1<sub>C</sub>), 5.636 (s, 1 H, CHPh), 5.534 (dd, 1 H, H- $^{2}$ <sub>D</sub>), 5.415 (d, 1 H, H- $^{1}$ <sub>D</sub>), 5.094 (t, 1 H, H- $^{4}$ <sub>D</sub>), 5.023 (d, 1 H,  $J_{1,2}$ 3.8 Hz, H-1<sub>B</sub>), 3.294 (s, 3 H,  $CH_3O$ ), 1.683 (s, 3 H,  $CH_3CO$ ), 1.344 and 0.651 (2 d, 2 H,  $J_{56} \sim 6.0$  Hz, H-6<sub>A</sub>, 6<sub>D</sub>); <sup>13</sup>C,  $\delta$  169.6 (C=O of Ac), 166.8 and 165.8 (C=O of Bz), 138.1–126.0 (aromatic), 100.5 (CHPh), 97.82 and 97.80 (C-1<sub>A</sub>,1<sub>D</sub>), 97.0 (C-1<sub>C</sub>), 92.9 (C-1<sub>B</sub>), 83.1 (C-4<sub>B</sub>), 76.0, 74.4, 73.6, 73.5, and 72.3 (CH<sub>2</sub> of Bn), 70.5 and 68.7  $(C-6_B,6_C)$ , 54.8  $(CH_3O)$ , 51.6  $(C-2_B)$ , 23.0  $(CH_3CO)$ , 18.0 and 17.4  $(C-6_A,6_B)$ . FABMS: m/z 1436 [(M + H)<sup>+</sup>]. Anal. Calcd for  $C_{83}H_{84}NO_{21}$ : C, 69.39; H, 6.24; N, 0.98. Found: C, 69.41; H, 6.34; N, 0.91.

2-(Trimethylsilyl)ethyl O-(3,4,6-tri-O-benzoyl-α-D-galactopyranosyl)-(1  $\rightarrow$  3)-O-(2-azido-4,6-O-benzylidene-2-deoxy-α-D-glucopyranosyl-(1  $\rightarrow$  3)-2,4-di-O-benzoyl-α-L-rhamnopyranoside (33).—A mixture of 32 (ref 6) (1.0 g, 0.75 mmol), 2,3-dichloro-5,6-dicyano-1,4-benzoquinone (220 mg, 0.97 mmol), CH<sub>2</sub>Cl<sub>2</sub> (100 mL), and H<sub>2</sub>O (5 mL) was stirred at 25°C for 6 h. More 2,3-dichloro-5,6-dicyano-1,4-benzoquinone (290 mg, 1.27 mmol) was added and stirring was continued for another 24 h. The mixture was extracted with aq NaHCO<sub>3</sub> and H<sub>2</sub>O. The organic phase was concentrated. Column chromatography (2:1 hexane-EtOAc) of the residue afforded 33 as an amorphous solid (860 mg, 94%);  $[\alpha]_D$  + 124° (c 0.2). NMR (CDCl<sub>3</sub>):  $^1$ H, δ 8.18-7.06 (aromatic protons), 5.856 (dd, 1 H,  $J_{3,4}$  3.2 Hz  $J_{4,5}$  1.9 Hz, H-4<sub>C</sub>), 5.625 (dd, 1 H,  $J_{1,2}$  1.3,  $J_{2,3}$  3.3 Hz, H-2<sub>A</sub>), 5.585 (t, 1 H,  $J_{3,4}$  =  $J_{4,5}$  = 10 Hz, H-4<sub>A</sub>), 5.520 (dd, 1 H,  $J_{2,3}$  10.5,  $J_{2,3}$  3.3 Hz, H-3<sub>C</sub>), 5.347 (s, 1 H, HCPh), 5.250

(d, 1 H,  $J_{1,2}$  3.7 Hz, H-1<sub>C</sub>), 5.167 (d, 1 H,  $J_{1,2}$  3.6 Hz, H-1<sub>B</sub>), 4.998 (d, 1 H,  $J_{1,2}$  1.7 Hz, H-1<sub>A</sub>), 3.106 (dd, 1 H, H-2<sub>B</sub>), 1.350 (d, 3 H, H-6<sub>A</sub>), 1.1–0.9 (m, 2 H, C $H_2$ Si), and 0.084 [s, 9 H, SiMe<sub>3</sub>]; <sup>13</sup>C,  $\delta$  166.0–165.3 (C=O), 136.2–123.7 (aromatic), 101.4 (CHPh), 101.3 (C-1<sub>A</sub>), 97.1 (C-1<sub>B</sub>), 95.3 (C-1<sub>A</sub>), 81.0 (C-4<sub>B</sub>), 68.3 (C-6<sub>B</sub>), 65.8 (OC+1<sub>2</sub>CH<sub>2</sub>), 61.7 (C-6<sub>C</sub>), 18.0 (C+1<sub>2</sub>Si), 17.7 (C-6<sub>A</sub>), and –1.3 (SiMe<sub>3</sub>). FABMS: m/z 1222 [(M + H)<sup>+</sup>] and 1196 [(M + H<sub>3</sub> – N<sub>3</sub>)<sup>+</sup>]. Anal. Calcd for C<sub>65</sub>H<sub>67</sub>N<sub>3</sub>O<sub>19</sub>Si: C, 63.87; H, 5.52; N, 3.44. Found: C, 63.74; H, 5.58; N, 3.41.

2-(Trimethylsilyl)ethyl  $O-(2,3,4-tri-O-benzoyl-\alpha-L-rhamnopyranosyl)-(1 \rightarrow 2)-O-$ (3,4.6-tri-O-benzovl- $\alpha$ -D-galactopyranosyl)- $(1 \rightarrow 3)$ -O-(2-azido-4.6-O-benzyldine-2 $deoxy-\alpha-D-glucopyranosyl$ )- $(1 \rightarrow 3)-2,4-di$ -O-benzoyl- $\alpha$ -L-rhamnopyranoside (35).—A mixture of 33 (700 mg, 0.57 mmol), 2,3,4-tri-O-benzoyl- $\alpha$ -L-rhamnopyranosyl bromide [26] (34, 500 mg, 0.93 mmol), 2,6-di-tert-butyl-4-methylpyridine (190 mg, 0.93 mmol), 4A molecular sieves (0.5 g), and dry (CH<sub>2</sub>Cl)<sub>2</sub> (20 mL) was stirred for 1 h then cooled to  $-20^{\circ}$ C. Silver trifluoromethanesulfonate (250 mg, 1.13 mmol) was added. The stirred mixture was allowed to reach 20°C in 4 h. Ice-cold, aq NaHCO<sub>3</sub> was added and the mixture was filtered. The organic phase was separated and concentrated. Column chromatography (3:1 hexane-EtOAc) of the residue afforded 35 as an amorphous solid (905 mg, 94%);  $[\alpha]_D + 149^\circ$  (c 0.6). NMR (CDCl<sub>3</sub>):  ${}^{1}$ H,  $\delta$  8.62–6.82 (aromatic), 5.983 (dd, 1 H,  $J_{3,4}$  3.3,  $J_{4,5}$  2.1 Hz, H-4<sub>C</sub>), 5.858 (dd, 1 H,  $J_{2,3}$  9.7 Hz, H-3<sub>C</sub>), 5.738 (dd, 1 H, H-3<sub>D</sub>), 5.723 (d, 1 H,  $J_{1,2}$  4.2 Hz,  $H-1_{C}$ ), 5.686 (dd, 1 H,  $J_{1,2}$  1.8,  $J_{2,3}$  Hz, 3.2 Hz,  $H-2_{A}$ ), 5.598 (t, 1 H,  $J_{3,4} = J_{4,5} = 9.8$ Hz, H-4<sub>A</sub>), 5.523 (s, 1 H, HCPh), 5.439 (dd, 1 H, H-2<sub>D</sub>), 5.005 (d,  $J_{1,2}$  1.7 Hz,  $\text{H-1}_{\text{A}}$ ), 3.484 (dd, 1 H,  $J_{2.3}$  9.9 Hz,  $\text{H-2}_{\text{B}}$ ), 1.342 and 0.611 (2 d, 6 H,  $\text{H-6}_{\text{A}}$ ,  $6_{\text{D}}$ ), and 0.085 [s, 9 H, SiMe<sub>3</sub>];  $^{13}$ C,  $\delta$  136.7–126.1 (aromatic carbons), 101.1 (CHPh), 97.3, 97.2, and 97.1 ( $C-1_A,1_C,1_D$ ), 94.8 ( $C-1_B$ ), 82.5 ( $C-4_B$ ), 68.2 ( $C-6_B$ ), 65.7  $(OCH_2CH_2)$ , 61.9  $(C-6_C)$ , 17.9  $(CH_2Si)$ , 17.6 and 17.2  $(C-6_A,6_D)$  and -1.3 (SiMe<sub>3</sub>). Anal. Calcd for  $C_{92}H_{89}N_3O_{26}Si$ : C, 65.74; H, 5.34; N, 2.50. Found: C, 65.52; H, 5.39; N, 2.44.

2-(Trimethylsilyl)ethyl  $O-(2,3,4-tri-O-benzoyl-\alpha-L-rhamnopyranosyl)-(1 \rightarrow 2)-O-$ (3,4,6-tri-O-benzoyl- $\alpha$ -D-galactopyranosyl)- $(1 \rightarrow 3)$ -O-(4,6-di-O-acetyl-2-azido-2deoxy- $\alpha$ -D-glucopyranosyl)- $(1 \rightarrow 3)$ -2,4-di-O-benzoyl- $\alpha$ -L-rhamnopyranoside (36).— To a stirred solution of 35 (830 mg, 0.49 mmol) in AcOH (10 mL) was added H<sub>2</sub>O at 70°C. After 5 h the mixture was concentrated. Toluene was added and evaporated from the residue. To a solution of the syrupy residue in pyridine (5 mL) were added Ac<sub>2</sub>O (5 mL) and a catalytic amount of 4-dimethylaminopyridine. After 5 h the mixture was processed as usual, followed by column chromatographic purification (3:1 hexane-EtOAc) to give 36 as an amorphous solid (760 mg, 92%);  $[\alpha]_{\rm D}$  + 160° (c 0.5). NMR (CDCl<sub>3</sub>): <sup>1</sup>H,  $\delta$  8.14–7.04 (aromatic), 5.955 (dd, 1 H,  $J_{3.4}$ 3.4,  $J_{4.5}$  1.6 Hz, H-4<sub>C</sub>), 5.744 (dd, 1 H,  $J_{2.3}$  10.6 Hz, H-3<sub>C</sub>), 5.720 (dd, 1 H, H-3<sub>D</sub>), 5.648 (dd, 1 H,  $J_{1,2}$  1.6,  $J_{2,3}$  3.5 Hz, H-2<sub>A</sub>), 5.539 and 5.532 (2 t, 2 H, H-4<sub>A</sub>, 4<sub>D</sub>), 5.307 (d, 1 H,  $J_{1,2}$  3.5 Hz, H-1<sub>B</sub>), 4.980 (d, 1 H, H-1<sub>A</sub>), 3.553 (dd, 1 H,  $J_{2,3}$  10.0 Hz,  $H-2_B$ ), 1.998 and 1.809 (2 s, 6 H, 2 C $H_3$ CO), 1.341 and 1.239 (2 d, 6 H,  $H-6_A$ ,  $6_D$ ), and 0.005 [s, 9 H, SiMe<sub>3</sub>];  $^{13}$ C,  $\delta$  170.8 and 169.2 (C=O of Ac), 166.0, 165.9, 165.8, 165.3, 164.9 (2 C), 164.8, and 164.6 (C=O of Bz), 133.4-128.1 (aromatic carbons), 98.8, 97.2, and 97.0 (C-1<sub>A</sub>,1<sub>C</sub>,1<sub>D</sub>), 93.9 (C-1<sub>B</sub>), 65.8 (OCH<sub>2</sub>CH<sub>2</sub>), 61.9 and 61.1

(C-6<sub>B</sub>, 6<sub>C</sub>), 20.6 (*C*H<sub>3</sub>CO), 18.1, 17.7, and 17.6 (*C*H<sub>2</sub>Si, C-6<sub>A</sub>, 6<sub>D</sub>), and -1.4 (SiMe<sub>3</sub>). FABMS: m/z 1651 [( $^{12}C_{88}^{13}CH_{89}N_3O_{28}Si - N_2 + H_3$ )<sup>+</sup>], 1650 [( $^{12}C_{89}H_{89}N_3O_{28}Si - N_2 + H_3$ )<sup>+</sup>], 1559 [( $^{12}C_{89}^{13}CH_{89}N_3O_{28}Si + H - SEOH$ )<sup>+</sup>], and 1558 [( $^{12}C_{89}H_{89}N_3O_{28}Si + H - SEOH$ )<sup>+</sup>]. Anal. Calcd for  $C_{89}H_{89}N_3O_{28}Si + C_{89}N_3O_{28}Si + C_{89}N_3$ 

O-2,3,4-Tri-O-benzoyl- $\alpha$ -L-rhamnopyranosyl)- $(1 \rightarrow 2)$ -O-(3,4,6-tri-O-benzoyl- $\alpha$ -Dgalactopyranosyl)- $(1 \rightarrow 3)$ -O-(4,6-di-O-acetyl-2-azido-2-deoxy- $\alpha$ -D-glucopyranosyl)- $(1 \rightarrow 3)$ -2,4-di-O-benzoyl-L-rhamnopyranose (37).—A solution of 36 (1.6 g, 0.95) mmol) in 1:4 CH<sub>2</sub>Cl<sub>2</sub>—CF<sub>3</sub>CO<sub>2</sub>H (40 mL) was kept at 25°C for 5 h. Toluene (3 × 5 mL) was added and the solution was concentrated under vacuum. Purification by column chromatography (3:2 hexane-EtOAc) afforded 37 as an amorphous solid (1.3 g, 87%);  $[\alpha]_D$  +178° (c 0.3). NMR (CDCl<sub>3</sub>): <sup>1</sup>H,  $\delta$  8.14–7.04 (aromatic), 5.963 (dd, 1 H, H- $^4$ C), 5.783 (dd, 1 H,  $J_{1,2}$  3.7,  $J_{2,3}$  10.2 Hz, H- $^3$ C), 5.723 (dd, 1 H, H-3<sub>D</sub>), 5.551 and 5.539 (2 t, 2 H, H-4<sub>A</sub>,  $\stackrel{7}{4}_{D}$ ), 5.438 (dd, 1 H,  $J_{1,2}$  1.7,  $J_{2,3}$  3.5 Hz, H-2<sub>A</sub>), 5.399 (d, 1 H,  $J_{1,2}$  3.5 Hz, H-1<sub>B</sub>), 2.010 and 1.816 (2 s, 6 H, 2  $CH_3CO$ ), 1.338 and 1.245 (2 d, 6 H,  $H_3CO$ ); <sup>13</sup>C,  $\delta$  170.8 and 169.2 (C=O of Ac), 165.9-164.6 (C=O of Bz), 133.5-128.1 (aromatic carbons), 98.7, 97.1, 93.8, and 92.4  $(C-1_A, 1_B, 1_C, 1_D)$ , 61.2 and 60.4  $(C-6_B, 6_C)$ , 20.65 and 20.61  $(CH_3CO)$ , 17.7 and 17.6  $(C-6_A, 6_D)$ . FABMS: m/z 1558  $[(M + H - H_2O)^+]$  and 1550  $[(M - N_2 + H_3)^+]$ . Anal. Calcd for C<sub>84</sub>H<sub>77</sub>N<sub>3</sub>O<sub>28</sub>: C, 64.00; H, 4.92; N, 2.66. Found: C, 63.81; H, 5.01; N, 2.57.

 $O-(2,3,4-Tri-O-benzoyl-\alpha-L-rhamnopyranosyl)-(1 \rightarrow 2)-O-(3,4,6-tri-O-benzoyl-\alpha-L-rhamnopyranosyl)$ D-galactopyranosyl)- $(1 \rightarrow 3)$ -O-(4.6-di-O-acetyl-2-azido-2-deoxy- $\alpha$ -D-glucopyranosyl)- $(1 \rightarrow 3)$ -2,4-di-O-benzoyl- $\alpha$ -L-rhamnopyranosyl trichloroacetimidate (38).—To a stirred solution of 37 (1.5 g, 0.95 mmol) in  $CH_2Cl_2$  (15 mL) were added at  $-20^{\circ}C$ trichloroacetonitrile (3.0 mL, 30 mmol) and 1,8-diazabicyclo[5.4.0]undec-7-ene (80  $\mu$ L, 0.53 mmol). The solution was stirred at  $-20^{\circ}$ C for 1 h and was then allowed to reach ~ 20°C in 1 h. Removal of the volatiles followed by purification by column chromatography (2:1 hexane-EtOAc) gave 38 as an amorphous solid (1.54 g, 94%);  $[\alpha]_D + 151^\circ$  (c 0.7). NMR (CDCl<sub>3</sub>): <sup>1</sup>H,  $\delta$  8.823 (s, 1 H, HN=C), 8.16–7.04 (aromatic protons), 6.448 (d, 1 H,  $J_{1,2}$  1.9 Hz, H-1<sub>A</sub>), 5.958 (dd, 1 H,  $J_{3,4}$  3.3,  $J_{4,5}$ 2.0 Hz,  $H-4_{\text{C}}$ ),  $5.888 \text{ (dd, 1 H, H-2}_{\text{A}}$ ),  $5.780 \text{ (dd, 1 H, } J_{2.3} \text{ 10.7 Hz}, H-3_{\text{C}}$ ),  $5.725 \text{ (dd, 1 H, } J_{2.3} \text{ 10.7 Hz}$ 1 H,  $J_{2,3}$  3.4,  $J_{3,4}$  9.9 Hz, H-3<sub>D</sub>), 5.636 (t, 1 H,  $J_{3,4} = J_{4,5} = 9.7$  Hz, H-4<sub>A</sub>), 5.536 (t, 1 H,  $J_{3,4} = J_{4,5} = 9.7$  Hz, H-4<sub>D</sub>), 5.436 (dd, 1 H, H-2<sub>D</sub>), 5.311 (d, 1 H,  $J_{1,2}$  3.5 Hz,  $H-1_C$ ), 5.262 (d, 1 H,  $J_{1,2}$  3.5 Hz,  $H-1_B$ ), 5.205 (d, 1 H,  $J_{1,2}$  1.7 Hz,  $H-1_D$ ), 5.187 (dd, 1 H, H-4<sub>B</sub>), 3.611 (dd, 1 H,  $J_{2,3}$  10 Hz, H-2<sub>B</sub>), 2.008 and 1.814 (3 s, 6 H, 3 C $H_3$ CO), 1.390 (d, 3 H,  $J_{5,6}$  6.2 Hz, H-6<sub>A</sub>), and 1.225 (d, 3 H,  $J_{5,6}$  6.1 Hz, H-6<sub>D</sub>). FABMS: m/z 1559 [( $^{12}C_{85}^{13}CH_{77}CI_3N_4O_{28} + H - C_2H_2CI_3NO$ )<sup>+</sup>] 1558  $[(C_{86}H_{77}Cl_3N_4O_{28} + H - C_2H_2Cl_3NO)^+], 1533 [(1559 - N_2 + H_2)^+], and 1532$  $[(1558 - N_2 + H_2)^+]$ . Anal. Calcd for  $C_{86}H_{77}Cl_3N_4O_{28}$ : C, 60.02; H, 4.51; N, 3.26; Cl, 6.18. Found: C, 59.81; H, 4.54; N, 3.20; Cl, 6.28.

Methyl  $O-(2,3,4-tri-O-benzoyl-\alpha-L-rhamnopyranosyl)-(1 \rightarrow 2)-O-(3,4,6-tri-O-benzoyl-\alpha-D-galactopyranosyl)-(1 \rightarrow 3)-O-(4,6-di-O-acetyl-2-azido-2-deoxy-\alpha-D-gluco-pyranosyl)-(1 \rightarrow 3)-O-(2,4-di-O-benzoyl-\alpha-L-rhamnopyranosyl)-(1 \rightarrow 3)-O-(2,4-di-O-benzoyl-\alpha-L-rhamnopyranosyl)-(1 \rightarrow 2)-O-(3,4,6-tri-O-benzyl-\alpha-D-galactopyranosyl)-$ 

 $(1 \rightarrow 3)$ -O-(2-acetamido-4,6-O-benzylidene-2-deoxy- $\alpha$ -D-glucopyranosyl)- $(1 \rightarrow 3)$ -2,4di-O-benzyl-α-L-rhamnopyranoside (39).—A mixture of 31 (250 mg, 0.174 mmol), 38 (200 mg, 0.116 mmol), and 4A molecular sieves (1.3 g) in dry CH<sub>2</sub>Cl<sub>2</sub> (8 mL) was stirred for 1 h then cooled to  $-20^{\circ}$ C. Boron trifluoride ethereate (20  $\mu$ L) was added, then the mixture was allowed to reach  $\sim -5-0^{\circ}$ C in 4 h. Triethylamine ( $\sim 200 \mu L$ ) was added. The mixture was filtered. The volatiles were removed under reduced pressure. Column chromatography (3:2 hexane-EtOAc) afforded **39** as an amorphous solid (252 mg, 69%);  $[\alpha]_D + 120^\circ$  (c 0.7). NMR (CDCl<sub>3</sub>): <sup>1</sup>H,  $\delta$ 6.202 (d, 1 H,  $J_{NH-H-2}$  9.5 Hz, NH), 5.902 (dd, 1 H,  $J_{3,4}$  3.1,  $J_{4,5}$  1.1 Hz, H-4<sub>G</sub>), 5.746-5.670 (m, 3 H, H- $^{2}$ <sub>D</sub>, $^{3}$ <sub>G</sub>, $^{3}$ <sub>H</sub>), 5.641 (s, 1 H, CHPh), 5.515 (t, 1 H, H- $^{4}$ <sub>Rha</sub>), 5.412 (d, 1 H,  $J_{1,2}$  1.5 Hz, H-1<sub>D</sub>), 5.356 and 5.346 (2 t, 2 H, 2 H-4<sub>Rha</sub>), 5.081 (dd, 1 H, H-4<sub>F</sub>), 5.019 (d, 1 H,  $J_{1,2}$  3.7 Hz, H-1<sub>B</sub>), 3.293 (s, 3 H, C $H_3$ O), 1.963 (s, 3 H, CH<sub>3</sub>CON), 1.752 and 1.667 (2 s, 6 H, 2 CH<sub>3</sub>CO), 1.335, 1.216, 1.186, and 0.656 (4 d, 12 H,  $J_{5.6} \sim 6.2$  Hz, H-6<sub>A</sub>,6<sub>D</sub>,6<sub>E</sub>,6<sub>H</sub>); <sup>13</sup>C,  $\delta$  170.5, 169.4, and 168.8 (C=O of Ac), 165.6-164.3 (C=O of Bz), 137.8-125.7 (aromatic), 100.5 (CHPh), 98.9, 98.6, 97.6, and 97.4 (C- $1_A$ , $1_D$ , $1_E$ , $1_H$ ), 96.8 and 96.7 (C- $1_C$ , $1_G$ ), 93.3 and 92.7 (C- $1_B$ ,  $1_E$ ), 82.7  $(C-4_B)$ , 75.7, 74.1, 73.4, 73.3, 72.0, 70.3, and 68.4  $(CH_2 \text{ of Bn and } C-6_B, 6_C)$ , 54.6 (CH<sub>3</sub>O), 51.3 (C-2<sub>R</sub>), 22.8 (CH<sub>3</sub>CON), 20.35 and 20.31 (CH<sub>3</sub>COO), 17.8, 17.3, and 17.2 (2 C) (C- $6_A$ , $6_D$ , $6_E$ , $6_F$ ). FABMS: m/z 2994.028 [(C<sub>167</sub>H<sub>164</sub>N<sub>4</sub>O<sub>48</sub> + H)<sup>+</sup>]. Anal. Calcd for C<sub>167</sub>H<sub>164</sub>N<sub>4</sub>O<sub>48</sub>: C, 66.97; H, 5.52; N, 1.87. Found: C, 65.20; H, 5.43; N, 1.92.

Methyl  $O-\alpha-1$ -rhamnopyranosyl- $(1 \rightarrow 2)$ - $O-\alpha-1$ -galactopyranosyl- $(1 \rightarrow 3)$ -O-(2-1)acetamido-2-deoxy- $\alpha$ -D-glucopyranosyl)- $(1 \rightarrow 3)$ -O- $\alpha$ -L-rhamnopyranosyl- $(1 \rightarrow 3)$ -O- $\alpha$ -L-rhamnopyranosyl- $(1 \rightarrow 2)$ -O- $\alpha$ -D-galactopyranosyl- $(1 \rightarrow 3)$ -O-(2-acetamido-2 $deoxy-\alpha-D-glucopyranosyl$ )- $(1 \rightarrow 3)-\alpha-L-rhamnopyranoside$  (11).—To a solution of 39 (240 mg, 0.08 mmol) in 1,2-dimethoxyethane (0.5 mL) was added a solution of NiCl<sub>2</sub> · 6 H<sub>2</sub>O (0.3 g, 1.26 mmol) and H<sub>3</sub>BO<sub>3</sub> (0.15 g, 2.4 mmol) in EtOH (10 mL). To this solution was added, under stirring at 25°C, a 1% solution of NaBH<sub>4</sub> in EtOH ( $\sim 40$  mL) during 1 h. The mixture was cooled to 0°C, then treated with Ac<sub>2</sub>O (2 mL). After 10 min the volatiles were removed under diminished pressure. The residue was equilibrated between CHCl<sub>3</sub> and H<sub>2</sub>O. The CHCl<sub>3</sub> phase was concentrated. Column chromatography of the residue (1:1 hexane-EtOAc) afforded **40** as a solid glass (105 mg, 44%). NMR (CDCl<sub>3</sub>):  $^{1}$ H,  $\delta$  6.580 and 6.290 (2 d, 2 H,  $J_{NH-H-2}$  9.5 Hz,  $NH_B$ ,  $NH_F$ ), 5.935 (br d, 1 H,  $H-4_G$ ), 5.587 (s, 1 H, CHPh), 3.293 (s, 3 H,  $CH_3O$ ), 1.946 and 1.927 (2 s, 6 H, 2  $CH_3CON$ ), 1.680 and 1.634 (2 s, 6 H, 2 C $H_3$ CO), 1.337, 1.160, 1.000, and 0.648 (4 d, 12 H,  $J_{5,6} \sim 6.1$  Hz,  $H-6_A, 6_D, 6_E, 6_H$ ); <sup>13</sup>C,  $\delta$  170.8, 170.0, 169.8, and 168.9 (C=O of Ac), 165.9–164.4 (C=O of Bz), 100.8 (CHPh), 99.0 (2 C), 98.6, 97.8 (2 C), 97.6, 97.4, and 93.0  $(C-1_A, 1_B, 1_C, 1_D, 1_E, 1_E, 1_G, 1_H)$ , 83.0  $(C-4_B)$ , 54.8  $(CH_3O)$ , 51.5  $(C-2_B)$ , 23.0 and 22.5  $(CH_3CON)$ , 20.8 and 20.6  $(CH_3COO)$ , 18.0, 17.5 (2 C), and 17.3  $(C-6_A, 6_D, 6_E,$  $6_{\rm H}$ ). FABMS: m/z 3010.059 [( $C_{169}H_{168}N_2O_{49}+H$ )<sup>+</sup>]. A solution of 40 in MeOH was treated with NaOMe until the pH of the solution reached  $\sim 12-13$  at 25°C. After the 12 h the solution was neutralized with Dowex  $50 \times 2$  (H<sup>+</sup>) and filtered, then the volatiles were removed. The residue was purified by column chromatography, using 3:1 EtOAc-CH<sub>3</sub>OH as eluant. The resulting material was hydrogenolyzed in 10:1 EtOH–AcOH on 10% Pd–C (Degussa Type E101NE/W) at 25°C for 24 h, under atmospheric pressure. The usual work-up followed by gel filtration through Biogel P-2 using 0.02 M pyridinium acetate as eluant afforded 11 as an amorphous solid;  $[\alpha]_D$  +77° (c 0.3, H<sub>2</sub>O). NMR (D<sub>2</sub>O): <sup>1</sup>H,  $\delta$  5.600 (d, 2 H,  $J_{1,2}$  3.5 Hz, H-1<sub>C</sub>,1<sub>G</sub>), 5.108 (d, 1 H,  $J_{1,2}$  1.5 Hz, H-1<sub>E</sub>), 5.081 (d, 1 H,  $J_{1,2}$  1.8 Hz, H-1<sub>H</sub>), 5.057 (d, 1 H,  $J_{1,2}$  1.5 Hz, H-1<sub>D</sub>), 5.042 (d, 1 H,  $J_{1,2}$  3.2 Hz, H-1<sub>F</sub>), 4.992 (d, 1 H,  $J_{1,2}$  3.3 Hz, H-1<sub>B</sub>), 4.714 (d, 1 H,  $J_{1,2}$  1.6 Hz, H-1<sub>A</sub>), 3.394 (s, 3 H, CH<sub>3</sub>O), 2.060 and 2.052 (2 s, 6 H, 2 CH<sub>3</sub>CO), 1.350–1.290 (4 d, 12 H, H-6<sub>A</sub>,6<sub>D</sub>,6<sub>E</sub>,6<sub>H</sub>); <sup>13</sup>C,  $\delta$ 174.76 and 174.70, (C=O), 102.9, 102.2, 102.1, and 101.4 (C-1<sub>A</sub>, 1<sub>D</sub>, 1<sub>E</sub>, 1<sub>H</sub>), 98.4 (2 C) (C-1<sub>C</sub>,1<sub>G</sub>), 94.8 (2 C) (C-1<sub>B</sub>,1<sub>F</sub>), 78.8 (C-3<sub>D</sub>), 61.5 (2 C) (C-6<sub>C</sub>,6<sub>G</sub>), 60.76 and 60.71 (C-6<sub>B</sub>,6<sub>F</sub>), 55.5 (CH<sub>3</sub>O), 52.7 (C-2<sub>B</sub>, 2<sub>F</sub>), 22.8 (CH<sub>3</sub>CON), 17.6, 17.48, 17.42, and 17.36 (C-6<sub>A</sub>,6<sub>D</sub>,6<sub>E</sub>,6<sub>H</sub>). FABMS: m/z 1369 [(C<sub>53</sub>H<sub>90</sub>N<sub>2</sub>O<sub>37</sub> + Na)<sup>+</sup>] and 1347 [(C<sub>53</sub>H<sub>90</sub>N<sub>2</sub>O<sub>37</sub> + H)<sup>+</sup>].

# References

- [1] V. Pozsgay, C.P.J. Glaudemans, J.B. Robbins, and R. Schneerson, *Bioorg. Med. Chem. Lett.*, 2 (1992) 255-260.
- [2] J.B. Robbins, C. Chu, and R. Schneerson, Clin. Inf. Dis., 15 (1992) 346-362.
- [3] J.B. Robbins, C. Chu, D.C. Watson, S.C. Szu, E.M. Daniels, C.U. Lowe, and R. Schneerson, Rev. Inf. Dis., 13 (Suppl) 4 (1991) s362-s365.
- [4] V. Pozsgay, B. Coxon, and H. Yeh, Bioorg. Med. Chem., 1 (1993) 237-257.
- [5] V. Pozsgay and B. Coxon, Carbohydr. Res., 257 (1994) 189-215.
- [6] V. Pozsgay, C.P.J. Glaudemans, J.B. Robbins, and R. Schneerson, Carbohydr. Res., 244 (1993) 259-273.
- [7] V. Pozsgay, C.P.J. Glaudemans, J.B. Robbins, and R. Schneerson, Tetrahedron, 48 (1992) 10249– 10264.
- [8] B.A. Dmitriev, Yu. A. Knirel, N.K. Kochetkov, and I.L. Hofman, Eur. J. Biochem., 66 (1976) 559-566.
- [9] S. Sturm, B. Jann, K. Jann, P. Fortnagel, and K.N. Timmis, Microb. Pathog., 1 (1986) 307-324.
- [10] V. Pozsgay and B. Coxon, unpublished results.
- [11] V. Pavliak, E.M. Nashed, V. Pozsgay, P. Kovac, A. Karpas, C. Chu, R. Schneerson, J.B. Robbins, and C.P.J. Glaudemans, J. Biol. Chem., 268 (1993) 25797-25802.
- [12] H. Paulsen and V. Sinwell, Chem. Ber., 111 (1978) 869-878.
- [13] H. Lönn, Carbohydr. Res., 139 (1985) 105-113.
- [14] Y. Oikawa, T. Yoshioko, and O. Yonemitsu, Tetrahedron Lett., 23 (1982) 885-888.
- [15] C.A.A. van Boeckel and T. Beetz, Tetrahedron Lett., 24 (1983) 3775-3778.
- [16] P.J. Garegg and H. Hultberg, Carbohydr. Res., 72 (1979) 276-279.
- [17] H.-P. Wessel and D.R. Bundle, Carbohydr. Res., 124 (1983) 301-311.
- [18] V. Pozsgay, Carbohydr. Res., 235 (1992) 295-302.
- [19] H.-P. Wessel and D.R. Bundle, J. Chem. Soc., Perkin Trans. 1, (1985) 2251-2260.
- [20] V. Pavliak, P. Kovac, and C.P.J. Glaudemans, Carbohydr. Res., 229 (1991) 103-116.
- [21] H. Gross, I. Farkas, and R. Bognár, Z. Chem., 18 (1978) 201-210.
- [22] K. Bock and C. Pedersen, Acta Chem. Scand., Ser. B, 29 (1975) 258-264.
- [23] K. Jansson, Ph.D. Thesis, University of Lund, Sweden, 1991.
- [24] G. Magnusson, Trends Glycosci. Glycotechnol., 4 (1992) 358-367.
- [25] K.P.R. Kartha, M. Kiso, and A. Hasegawa, J. Carbohydr. Chem., 8 (1989) 675-679.
- [26] V. Pozsgay and H.J. Jennings, J. Org. Chem., 53 (1988) 4042-4052.
- [27] R.R. Schmidt and J. Michel, Angew. Chem. Int. Ed. Engl., 19 (1980) 731-732.

- [28] M. Numata, M. Sugimoto, K. Koike, and T. Ogawa, Carbohydr. Res., 163 (1987) 209-225.
- [29] V. Pozsgay and B. Coxon, in preparation.
- [30] A. Lipták, J. Imre, and P. Nánási, Carbohydr. Res., 92 (1981) 154-156.
- [31] Yu. E. Tsvetkov, L.V. Backinowsky, and N.K. Kochetkov, Carbohydr. Res., 193 (1989) 75-90.